

Tuning the membrane permeability of polymersome nanoreactors developed by aqueous emulsion polymerization-induced self-assembly

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Compartmentalization and Confinement in Nature







In all living organisms:

- Structural and functional organization on both cellular and subcellular levels (*i.e.* organelles)
- Selective communication and transport of energy, nutrients and signaling molecules
- Vital biological reactions occur in confined compartments
- Protection from external detrimental conditions



Transport of Molecules Across Cell/Organelle Membranes



Application of Membrane Diffusion (Glycolysis and Krebs Cycle for ATP production)



Two Main Pathways for Molecule Transport across Membranes:

1. Passive Transport:

Either <u>non-specific</u> or <u>specific</u> permeability Simple diffusion across semi-permeable membranes Carrier-mediated transport *via* channel membrane proteins

2. Active Transport:

Diffusion across the membrane requires the consumption of energy as fuel

Different diffusion/transport mechanisms allow for vital enzyme-catalyzed cascade reactions (e.g. cellular respiration) to occur selectively in confined environments

Structural and Functional Cell-Mimicry



Platform Design

Conventional Methodologies for Small Molecule Encapsulation



Current Opinion in Pharmacology

Liposomes & Polymersomes:

Self-assembled bilayer nanostructures

Minimal cell-mimicking synthetic analogues

Able to incorporate both hydrophilic and hydrophobic cargo Application as delivery vehicles, functional artificial organelles and catalytic nanoreactors

Polymersomes have superior physical and chemical stability

Preparation of polymersomes *via* conventional block copolymer self-assembly:

Laborious multi-step methodologies

Dilute conditions – low solids concentration (≤ 1% w/w) Require post-polymerization processing to target pure morphologies

Require the use of toxic organic solvents

Catalytic Nanoreactors & Artificial Organelles



Peters, et al. Angew. Chem. Int. Ed., 2014, 53, 146-150



Tanner, et al. Nano Lett., 2013, 13, 2875-2883



Common methodologies for permeability enhancement of nanoreactors toward substrates and catalysis products involve:

- Incorporation of channel-forming transmembrane proteins (e.g. OmpF, aquaporins)
- Stimuli-responsive membranes
- Inherently permeable membranes

Stimuli-Responsive Membranes



Polymerization-Induced Self-Assembly (PISA)



Tan, et al. Macromolecules, 2017, 50, 5798-5806

Aqueous Photo-PISA:

- Ambient temperatures compared to thermally initiated PISA (50 70 °C)
- Oxygen-tolerant processes (under certain conditions)
- One-pot non-disruptive encapsulation of INPs, drugs, fluorophores and enzymes into polymersomes
- **Biologically relevant applications**

Facile access to higher-order morphologies

Compatible with aqueous/organic media

Radical (RAFT, ATRP, NMP) and non-radical methodologies (ROMP, ROP)

Karagoz, et al. Polym. Chem., 2014, 5, 350-355 Canning, et al. Macromolecules, 2016, 49, 1985-2001

Functional Polymersome Nanoreactors Developed in the O'Reilly Group



Blackman, et al. ACS Macro Lett., 2017, 6, 1263-1267

7

Functional Polymersome Nanoreactors Developed in the O'Reilly Group



Reduced o-Dianisidine **Oxidised o-Dianisidine** (colorless) (red-brown) H₃CO H₃CO H₃CO [HRP] H₂N NH₂ OCH₃ OCH₃ OCH₃ H_2O_2 H_2O **OmpF** trimer KAN 2008 Vesicle VS Bilayer Inherently permeable **OmpF-functionalized HRP-loaded vesicles HRP-loaded vesicles** 1.0 30-- Intensity II -+HRP/-OmpF Volume Number Intensity +HRP/+OmpF 20 0.8 Z-ave = 411 ± 4 nm -HRP/-OmpF $PD = 0.12 \pm 0.02$ % AAbs492 nm 0.6 10 100 1000 10000 $D_{\rm h}$ (nm) 0.4 0.2 0.0 15 30 5 10 20 25 Time (min)

Membrane Protein Incorporation

Emerging Challenge

Can we fabricate catalytic polymersome nanoreactors with programmed size-selective permeability exploiting the advantages of aqueous photo-PISA?







Quantitative monomer conversion (> 98%) after 2 hours

Optical microscopy – Heterogeneous monomer-*in*-water emulsion (HPMA is water-miscible, GlyMA is water-immiscible) prior to photo-PISA with monomer droplets of 5-30 μ m



 Quantitative retention of pendant epoxide groups (99%) after photo-PISA at 37 °C

!!During thermally initiated PISA at elevated temperatures (60 - 100 °C), epoxides can undergo partial hydrolysis!!

- Increase in polymerization rate and onset of particle micellization after *ca*. 20 min (~17% monomer conversion)
- HPMA/GlyMA ratio between 3.0 4.0 indicates the formation of statistical copolymers

Kinetic Investigation:



Varlas, et al. Nanoscale, 2019, 11, 12643-12654

40



HRP-loaded Polymersomes:



Polymersome Membrane Modification Methodology:



Involves ring-opening of pendant PGlyMA epoxide groups using a series of primary amines and cross-linking diamines of varying hydrophobicity as nucleophiles





FT-IR – Complete disappearance of epoxide ring characteristic peaks at 849 and 909 cm⁻¹ indicating quantitative ring-opening

Permeability reduction due to thicker and more densely packed membranes as crosslinking reduces diffusivity



absorbance decrease of **45 ± 5%** absorbance decrease of **69 ± 4%**



Normalized transmittance

Wavenumber (cm⁻¹)





Occurrence of particle agglomeration in the case of longer aliphatic C_4DA and C_6DA cross-linkers due to the development of intervesicular interactions

FT-IR indicates quantitative ring-opening of epoxide groups

Permeability reduction due to thicker and more hydrophobic (less hydrated) membranes that hinder diffusion

HRP-catalysed assay

absorbance decrease ranging from **50%** to **61%** depending on membrane thickness increase



Other Hydrophobic Primary Amines:



Significant increase in membrane thickness in all cases that is dependent on the hydrophobicity of the modifier (NMA > BA > PXDA)



Kinetic colorimetric assay at fixed [substrate]:

~30% reduction in product formation for PXDA-crosslinked nanoreactors

>80% absorbance decrease was measured in the case of BA and NMA-functionalized polymersomes, showing nearcomplete permeability blockage **Detailed Permeability Investigation:**



Effect of [DMB] and determination of observable permeability Compared to K_m of free HRP (0.79 ± 0.06 mM): K_m^* of PGlyMA nanoreactors increased by *ca*. 30% K_m^* of PGlyMA+PXDA nanoreactors increased by *ca*. 60% K_m^* of PGlyMA+BA nanoreactors increased by *ca*. 75% K_m^* of PGlyMA+NMA nanoreactors increased by *ca*. 230%!!

Rate of substrate turnover was dependent of polymersome membrane thickness in a non-linear manner and trended based on the hydrophobicity of the utilized amine

Control Experiments:



Effects of ring-opening conditions on HRP activity:





A series of control experiments show no loss of enzyme activity or enzyme modification after photo-PISA and under applied ring-opening conditions

Summary

- Aqueous photo-PISA is a facile methodology to prepare epoxy-functionalized enzyme-loaded polymersome nanoreactors in one-pot
- Membrane modification is readily feasible using as series of amines as nucleophiles for ringopening of pendant epoxide groups under mild reaction conditions
- Membrane modification resulted in increased thickness and reduced porosity/permeability relative to the non-functionalized particles
- Membrane thickness increase and blocking effect were more prominent as the hydrophobicity of utilized nucleophile increased
- Small molecule diffusion across nanoreactor membranes can be controlled by following our two-step methodology and by using appropriately selected amines





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